#### LIMITATIONS OF THE ASSAY

- 1. The binding activity of the antibodies and the activity of the enzyme used are temperature-dependent. It is therefore recommended using a thermostat in all three incubation steps. The higher the room temperature (+18°C to +25°C) during the incubation steps, the greater will be the absorbance values. Corresponding variations apply also to the incubation times. However, the calibrator is subject to the same influences, with the result that such variations will be largely compensated in the calculation of the result.
- 2. For duplicate determination the mean of the two values should be taken. If the two values deviate substantially from one another, retesting the samples is recommended.
- 3. For diagnosis, the clinical picture of the patient always needs to be taken into account along with the serological findings.

#### **BIBLIOGRAPHY**

- 1. Reichlin, M. Current perspectives on serological reactions in SLE patients, Clin. Exp. Immunol. 44:1-10, 1981.
- 2. Gonz'alez C; Martin T; Arroyo T; Garc'ia-Isidoro M; Navajo JA; Gonz'alez-Buitrago JM. Comparison and variation of different methodologies for the detection of autoantibodies to nuclear antigens (ANA). J Clin Lab Anal 1997;11(6):388-92.
- Emlen W; O'Neill L Clinical significance of antinuclear antibodies: comparison of detection with immunofluorescence and enzyme-linked immunosorbent assays. Arthritis Rheum 1997;40(9):1612-8.
- 4. Rothschild, B.M., Jones, J.V., Chesney, C., Pifer, D., Thompson, L.D., James, K.K. and H. Badger. Relationship of clinical findings in Systemic Lupus Erythematosus to sero-reactivity. Arthritis and Rheumatism 26: 45-51, 1983.
- Spronk PE; Bootsma H; Horst G; Huitema MG; Limburg PC; Cohen Tervaert JW; Kallenberg CG. Antineutrophil cytoplasmic antibodies in systemic lupus erythematosus. Br J Rheumatol 1996;35(7):625-31.
- Venables, P.J.W., Erhardt, C.C. and R.N. Maini. Antibodies to extractable nuclear antigens in rheumatoid arthritis: relationship to vasculitis and circulating immune complexes. Clin. Exp. Immunol. 39:16, 1980.
- Welin Henriksson E; Hansson H; Karlsson-Parra A; Pettersson I. Autoantibody profiles in canine ANA-positive sera investigated by immunoblot and ELISA. Vet Immunol Immunopathol 1998;61(2-4):157-70.
- Koh WH; Dunphy J; Whyte J; Dixey J; McHugh NJ. Characterisation of anticytoplasmic antibodies and their clinical associations [see comments]. Ann Rheum Dis 1995;54(4):269-73.
- 9. Parveen S; Morshed SA; Nishioka M. High prevalence of antibodies to recombinant CENP-B in primary biliary cirrhosis: nuclear immunofluorescence patterns and ELISA reactivities. J Gastroenterol Hepatol 1995;10(4):438-45.
- Harmon, C.E. Anti-nuclear antibodies in autoimmune diseases; significance and pathogenecity. Medical Clinics of North America 69:547, 1985.

#### SYMBOL KEYS

Temperature Limitation	Consult Instructions for use	Date of Manufacture	LOT Batch Number / Lot Number
Manufacturer	IVD In vitro Diagnostic Medical Device	This side up	Contains sufficient
Use by	REF Catalogue Number	Do not reuse	for <n> tests</n>



# Zephyr Biomedicals

A Division of Tulip Diagnostics (P) Ltd. M 46-47, Phase III B, Verna Industrial Estate, Verna, Goa - 403 722, INDIA. end. Office: Gitaniali Tulip Block Dr Antonio Do Repo Bach Alto Santacru

Regd. Office: Gitanjali, Tulip Block, Dr. Antonio Do Rego Bagh, Alto Santacruz, Bambolim Complex P.O., Goa - 403 202, INDIA. Website: www.tulipgroup.com Email: sales@tulipgroup.com



Enzyme Linked Immunosorbent Assay (ELISA) for semi quantitative detection of anti-nuclear IgG antibodies (ANA) in Human Serum FOR IN VITRO DIAGNOSTIC USE ONLY Store at 2°C to 8°C

#### INTENDED USE

Qualisa<sup>™</sup> ANA- ELISA test system is an enzyme linked immunosorbent assay for semi quantitative detection of anti-nuclear IgG antibodies (ANA) in human serum. For in Vitro Diagnostic Use only.

#### INTRODUCTION

Anti-nuclear antibodies (ANA) are auto-antibodies that bind to several nuclear antigens including double stranded DNA (dsDNA), single-stranded DNA (SSDNA), ribonucleic-protein (RNP), Sm, SS-A and SS-B. ANA are frequently present in patients with systemic lupus erythematosus (SLE) and less commonly in other autoimmune diseases such as Rheumatoid arthritis, Collagen vascular diseases, chronic liver diseases and systemic sclerosis (scleroderma). The assay is used as a screening procedure for autoimmune diseases.

#### PRINCIPLE

The Qualisa<sup>™</sup> ANA test kit is an enzyme linked immunosorbent assay. Diluted serum is added to microwells coated with purified nuclear antigens. If ANA IgG specific antibodies are present in diluted serum, they bind to the microwells coated with purified nuclear antigen. Washing of the microwells removes all unbound materials. Enzyme Conjugate is added to the microwells to bind to the antibody-antigen complex if present. And washing of the microwells removes the excess of conjugate. A solution of TMB substrate is added to allow hydrolysis of the substrate by the enzyme. The reaction is stopped after specified time using stop solution. The intensity of the colour generated is proportional to the amount of IgG specific antibody in the sample.

### MATERIALS AND COMPONENTS

- A. Materials provided with the test kit:
- 1. Coated Microwells: Microwells coated with Nuclear antigen
- 2. Calibrator: Ready to use
- 3. Sample Diluent: Ready to use
- 4. Negative Control: Ready to use
- 5. Positive Control: Ready to use
- 6. Wash Buffer Concentrate (20X)
- 7. Enzyme Conjugate: Ready to use.
- 8. TMB Substrate: Ready to use
- 9. Stop Solution: Ready to use
- B. Materials required but not provided:
- 1) Precision pipettes: 10-100µl, 20-200µl, 100-1000µl
- 2) Disposable pipette tips
- Distilled water
- 4) Disposable Gloves
- 5) ELISA reader
- 6) ELISA washer

#### STORAGE AND STABILITY

- 1. **Qualisa<sup>™</sup> ANA** kit is stable at 2-8°C up to the expiry date printed on the label.
- 2. Coated Microwells should be used within one month upon opening the pouch provided that once opened, the pouch must be resealed to protect from moisture. If the colour of the desiccant has changed from blue to white at the time of opening the pouch, another coated Microwells pouch should be used.
- 3. Diluted Wash Buffer is stable up to one week when stored at 2-8°C.

#### **SAMPLE COLLECTION**

- 1. Collect Blood specimen by venipuncture according to standard procedure.
- 2. Only serum should be used.
- 3. Avoid grossly hemolytic, lipemic or turbid samples.
- 4. Preferably use fresh samples. However specimens can be stored upto 48 hour at 2-8°C for short duration.
- 5. For longer storage, specimens can be frozen at  $-20^{\circ}$ C. Thawed samples must be mixed prior to testing.
- 6. Do not heat inactivate before use.

- 7. Specimen containing precipitate or particulate matter should be clarified by centrifugation prior to use.
- 8. Specimen should be free from particulate matter and microbial contamination.

#### PRECAUTIONS

- (1) Bring all reagents and specimen to room temperature before use.
- (2) Do not pipette any material by mouth.
- (3) Do not eat, drink or smoke in the area where testing is done.
- (4) Use protective clothing and wear gloves when handling samples.
- (5) Use absorbent sheet to cover the working area.
- (6) Immediately clean up any spills with sodium hypochlorite.
- (7) All specimens and controls should be considered potentially infectious and discarded appropriately.
- (8) Neutralize acid containing waste before adding hypochlorite.
- (9) Do not use kit after the expiry date.
- (10) Do not mix components of one kit with another.
- (11) Always use new tip for each specimen and reagent.
- (12) Do not allow liquid from one well to mix with other wells.
- (13) Do not let the strips dry in between the steps.

## **REAGENT PREPARATION**

- 1. All reagents should be brought to room temperature (18-25°C) and mixed by gently inverting or swirling prior to use. Do not induce foaming.
- 2. Dilute wash buffer 20 times (for example add 5ml concentrated buffer to 95ml of distilled or deionized water). Mix well before use.

#### **TEST PROCEDURE**

- 1. Place the desired number of coated strips into the holder.
- Prepare 1:21 dilutions by adding 10µl of the test samples to 200µl of sample diluent. Mix well.
   Please Note: The Negative control, Positive control and the calibrator are ready to use, DO NOT dilute.
- Dispense 100µl of diluted serum samples, negative control, positive control and calibrator into the appropriate wells. For the reagent blank, dispense 100µl of sample diluent in A1 well position. Shake the holder to remove air bubbles from the liquid and mix well. Incubate at room temperature (18-25°C) for 20 minutes.
- 4. After incubation, empty the microtitre wells and wash the plate 3 times with **350µl** of diluted wash buffer. Strike the microtitre plate sharply onto the absorbent paper towel to remove all residual water droplets.
- 5. Dispense 100µl of Enzyme Conjugate to each well and incubate at room temperature (18-25°C) for 20 minutes.
- 6. After incubation, empty the microtitre wells and wash the plate 3 times with **350µl** of diluted wash buffer. Strike the microtitre plate sharply onto the absorbent paper towel to remove all residual water droplets.
- 7. Dispense 100µl of TMB Substrate into each well and incubate at room temperature (18-25°C) for 10 minutes in dark.
- 8. Stop the reaction by adding **100µl** of Stop Solution to each well. Gently mix for 10 seconds until the blue color completely changes to yellow.
- 9. Read the optical density at 450/630 nm with a microtiter plate reader within 15 minutes.

Add 100µl of sample diluent in A1 Microwell as blank

Add 100µl NC, PC, Calibrator and diluted samples into the respective Microwells

Apply plate sealer and incubate for 20 minutes at 18-25°C

Add 100µl Enzyme Conjugate in each well

Add 100µl Enzyme Conjugate in each well

Apply plate sealer and incubate for 20 minutes at 18-25°C

Apply plate sealer and incubate for 20 minutes at 18-25°C

Apply plate sealer and incubate for 20 minutes at 18-25°C

Apply plate sealer and incubate for 10 minutes at 18-25°C

Add 100µl Stop Solution in each well

Add 100µl Stop Solution in each well
Add 100µl Stop

#### **RUN CRITERIA**

- 1) The O.D of the Calibrator should be greater than 1.0 2) The ANA Index of the Negative control should be 0.9 or less.
- 3) The ANA index of the Positive control should be in the range as stated on the label.
- If any of these criteria are not met, the results are invalid and the test should be repeated.

#### **CALCULATION OF RESULTS**

1) To obtain the Cut off Value (COV): Multiply the OD of Calibrator by Calibrator Factor (CF) (which is lot specific & will be printed on the label of the calibrator vial). 2) Calculate the ANA Index of each determination by dividing the OD values of each sample by obtained OD of Cut off value.

## **For example**

If the Calibrator Factor (CF) on the label is 0.38 Calibrator mean OD = 2.001 Cut-off Value = 2.001 × 0.38 = 0.760 Patient sample OD = 1.594 ANA Index = 1.594/0.760 = 2.09 (Positive result) Patient Sample OD = 0.232 ANA Index = 0.232/0.760 = 0.30 (Negative result)

## INTERPRETATION OF THE RESULT

Negative: ANA Index of 0.90 or less. No detectable ANA IgG by ELISA. Equivocal: ANA Index of 0.91 - 1.1 are equivocal. Follow up testing is recommended if clinically indicated. Positive: ANA Index of 1.11 or above. Detectable ANA IgG by ELISA.

## PERFORMANCE CHARACTERISTICS

### 1) Sensitivity and Specificity

233 patient sera were tested by this ELISA and a reference ELISA method. 2 sera were positive and 220 sera were negative by both methods. The correlation between the two methods was 95.2%. The results are summarized as below:

Qualisa ANA IgG ELISA					
		+	+/-	-	Total
Reference ELISA Kit	+	2	2	6	10
	+/-	0	0	2	2
	-	1	0	220	221
Total		3	2	228	233

# 2) Precision:

Intra Assay study

Sample	No of replicates	Mean	Standard Deviation	Coefficient of variation (%)
1	16	1.79	0.08	4.60
2	16	1.36	0.043	3.21
3	16	0.37	0.042	11.4

## Inter Assay study

Sample	No of replicates	Mean	Standard Deviation	Coefficient of variation (%)
1	10	1.94	0.115	5.94
2	10	1.32	0.078	5.89
3	10	0.41	0.046	10.96

#### **IMPORTANT NOTE**

- 1. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.
- 2. It is recommended to use the multi-channel pipettes to avoid time effect. A full plate of 96 wells may be used if automated pipetting is available.

3. Duplication NC, PC, Calibrator & samples is not mandatory but may provide information on reproducibility & application errors.