

PERFORMANCE

The performance study has been evaluated on 3183 blood samples from 2734 donors, 331 patients, 65 newborns and 53 weak D blood samples. Each ABO/D test result including reverse typing, showed complete agreement with the analysis method of the reference laboratory in a university blood bank. Blood samples with weak D expression showed different reaction strength.

BIBLIOGRAPHY

1. Human Blood Groups by Geoff Daniels, 2nd Edition, Blackwell Science, Oxford 2002.
2. HMSO, Guidelines for the Blood Transfusion Services, 2nd Edition, 1993.
3. H. Malyska & D. Weiland, The Gel Test. Laboratory Medicine Vol. 25, No.2 February 1994, pg. 81-85.
4. M.C.Z. Novaretti et al. Comparison Of Tube And Gel Techniques For Antibody Identification. Immunohematology 2000;16:138-141.
5. D. Voak, New Developments in Blood Group Serology, Infusion Therapy Transfusion Medicine 1999;26:258-260.
6. Data on file: Tulip Diagnostics (P) Ltd.



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matrixTM
— GEL • SYSTEM

REF	10260024	10260048
Pack Size	24 Cards	48 Cards

MatrixTM ABO/Rho(D) Forward and Reverse Grouping Card with Autocontrol

SUMMARY

Human red blood cell antigen can be divided into four groups A, B, AB and O depending on the presence or absence of corresponding antigens on the red blood cells. The Anti-A, Anti-B and Anti-D reagents are used to detect the presence or absence of corresponding antigens on red blood cells.

Red blood cells used in Reverse grouping are of known ABO antigen, having the specificity to indicate the presence or absence of Anti-A and/or Anti-B, the result of which determines the reverse group.

Classification of blood groups must be based on both forward and reverse grouping. The MatrixTM Forward and Reverse Grouping Card with Autocontrol allows combined testing of ABO forward and reverse grouping as well as Rho (D) determination.

REAGENTS

MatrixTM Forward and Reverse Grouping Card with Autocontrol contains six microtubes prefilled with a gel in a suitable buffer containing Monoclonal Anti-A (Clone 11H5), Anti-B (Clone 6F9) and Anti-D (IgM) (VI-) (Clone P3x61 + TH-28) from microtube 1 to 3, microtube 4 to 6 contains neutral gel where microtube 4 (Ctrl) is negative control and microtube 5 and 6 (A, & B) serves for reverse grouping.

STORAGE AND STABILITY

Store MatrixTM gel cards in an upright position at 4-25°C. Do not freeze.

Avoid exposure of MatrixTM gel cards to direct sunlight or any heat source. The shelf life of MatrixTM gel cards is as per the expiry date mentioned on the label. Do not use beyond expiry date. Once the aluminium foil is removed from the microtube, it should be used immediately.

ADDITIONAL REAGENTS AND MATERIALS REQUIRED:

MatrixTM Diluent -2 LISS for preparation of red cell suspension. (Refer package insert before use). Gel card centrifuge (85g), Work station, Micropipette capable of delivering 5-50µl of specimen and Bottle top dispenser.

PRINCIPLE

As the MatrixTM gel card containing red blood cells is centrifuged under specific conditions, the red blood cells possessing the corresponding antigen will agglutinate in presence of the specific antibody and will be trapped in the gel column. The red blood cells, which do not react are not trapped in the gel column and get settled at the bottom of the microtube. The reactions are then read and graded according to their reactivity pattern.

SAMPLE COLLECTION

No special preparation of the patient is required prior to sample collection by approved techniques. For optimal results, freshly collected sample should be used. Anticoagulants like EDTA, CPD-A and Citrate can be used. Serum or plasma samples can be used.

Samples should be centrifuged at 1500g for 10 minutes to avoid fibrin residue which may interfere with results.

SAMPLE PREPARATION

For Forward Grouping

Prepare a 5% red blood cell suspension in MatrixTM Diluent-2 LISS as follows:

1. Bring the MatrixTM Diluent-2 LISS to room temperature before testing.
2. Dispense 0.5 ml of MatrixTM Diluent-2 LISS into a clean test tube.
3. Add 50µl of whole blood or 25µl of packed red cells to MatrixTM Diluent-2 LISS collected in test tube and mix gently.
4. Red blood cell suspension so obtained should be used for forward grouping.

For Reverse Grouping

Prepare a 0.8% red blood cell suspension in MatrixTM Diluent-2 LISS as follows:

1. Collect known A₁ and B cells from at least three donors and pool in respective test tubes labeled as A, & B.
2. Wash the cells with 0.9% saline till the supernatant is clear.
3. Dispense 1ml of MatrixTM Diluent-2 LISS into clean labeled test tubes (A, & B).

- Add 10µl of packed red blood cells (pooled and washed known A, & B cells) into respective test tubes and mix gently.
- Red blood cell suspensions so obtained should be used for reverse grouping.

TEST PROCEDURE

- Label the "Matrix™ Forward and Reverse Grouping Card with Autocontrol" with patient's name or identification number. Remove the aluminium foil carefully by pulling it backwards.
- Pipette 50µl of 0.8% known A, cell suspension to the microtube 5.
- Pipette 50µl of 0.8% known B cell suspension to the microtube 6.
- Pipette 50µl of patient's plasma or serum to the microtubes 5 and 6.
- Allow the card to incubate for 10 minutes at room temperature.
- Pipette 10µl of 5% patient's red cell suspension to the microtubes 1-4 (A-B-D-Ctrl), taking care to ensure that micropipette tip does not touch the microtube.
- Centrifuge the cards for 10 minutes in the gel card centrifuge.
- Retrieve the card from centrifuge, read and record the results.

INTERPRETATION OF RESULTS

The control microtube (Ctrl) must be negative to validate the forward grouping results. If not, then repeat the test after washing the patient's red blood cells with warm saline.

Positive reaction: Agglutinated red blood cells forming a clear line on the surface of gel column or agglutinates dispersed in the gel column.

Negative reaction: Non agglutinated red blood cells settle at the bottom of the microtube forming a compact button.

The reaction strength may be recorded as follows:

Strength of reaction	Comments
4+	Agglutinated red blood cells form a line at the top of the gel microtube.
3+	Most agglutinated red blood cells remain in the upper half of the gel microtube.
2+	Agglutinated red blood cells are observed throughout the length of the microtube. A small button of red blood cells may also be visible at the bottom of the gel microtube.
1+	Most agglutinated red blood cells remain in the lower half of the microtube. A button of cells may also be visible at the bottom of the gel microtube.
±	Most agglutinated red blood cells are in the lower third part of the gel microtube.
Negative	All the red blood cells pass through and form a compact button at the bottom of the gel microtube.
Mixed field agglutination	Agglutinated red blood cells form a line at the top of the gel and non-agglutinated red blood cells form a compact button at the bottom of the gel microtube.
H	Hemolysis of red blood cells

Expected reactivity pattern for ABO grouping:

Anti-A	Anti-B	Blood Group
± to 4+	Negative	A
Negative	± to 4+	B
± to 4+	± to 4+	AB
Negative	Negative	O

NOTE: Human red blood cells that show weak reaction with Anti-A and/or Anti-B probably indicate subgroups of A and/or B and further testing is recommended.

Expected reactivity pattern for Rho (D) typing:

Anti-D	Rho(D) Type
± to 4+	Rho (D) Positive
Negative	Rho (D) Negative

NOTE: Weak D/ Partial D type human red blood cells may give a weaker or negative reaction. Such cells should be retested for weak D confirmation with Matrix™ Coombs Anti-IgG card.

Reaction for reverse grouping:

A ₁	B	Blood group
± to 4+	Negative	B
Negative	± to 4+	A
± to 4+	± to 4+	O
Negative	Negative	AB

NOTE

- In vitro diagnostic reagent for laboratory and professional use only. Not for medicinal use.
- The Matrix™ gel cards contains sodium azide < 0.1% as preservative. Avoid contact with skin and mucosa. On disposal flush with large quantity of water.
- All Matrix™ gel cards should be centrifuged for one complete cycle (10 minutes) in gel card centrifuge before use.
- Visually inspect the Matrix™ gel cards before use.
- Matrix™ gel cards having bubble(s) entrapped within the gel can be centrifuged for two complete cycles in gel card centrifuge to remove the bubble, if bubbles are not removed the card should not be used.
- Matrix™ gel cards that exhibit any signs of drying (i.e. absence or reduced level of reagent buffer above the gel column), decreased volume of gel, cracked gel should not be used.
- Matrix™ gel cards with damaged aluminium foil seal should not be used.
- Freezing of Matrix™ gel cards or evaporation of gel or reagent buffer due to exposure to heat may lead to erroneous results.
- Fibrin or particulate matter if present in the sample may lead to erroneous results.
- Fibrin if present in the sample may trap red blood cells on top of gel column presenting a pink line. To avoid, samples should be well centrifuged at 1500g for 10 minutes before taking serum or plasma and RBCs should be washed if not collected properly in an anticoagulant.
- Use of red blood cells concentration/ volume and reagents other than those described may lead to erroneous results. Follow the instructions carefully.
- Aged or stored red blood cells may exhibit weaker reactivity than freshly collected cells.
- Old cell panels may give an unclear background with Matrix™ gel cards.
- Do not use hemolysed samples.
- Extreme turbidity or discoloration may indicate microbial contamination or denaturation of protein due to thermal damage. Such Matrix™ gel cards should be discarded.
- Contamination of reagents during usage may cause false positive or negative results.
- Red cell aggregation in the red cell suspension may interfere the passage.
- Aluminium foil seal of Matrix™ gel cards should be removed gently and carefully by pulling the foil seal backwards to avoid contamination of reagents from one microtube to another.
- Do not use lipemic, icteric and hyperproteic samples.
- To avoid contamination always use fresh tips before dispensing into each microtube.
- Matrix™ ContaVoid can be used to avoid contamination of reagents in microtubes while usage. For details refer pack insert of Matrix™ ContaVoid (Catalogue no.102770100)

REMARKS

- Known positive and negative control should be tested as per Good Laboratory Practices.
- ERYWELL (Catalogue no. 10253020) can be used as red blood cell preservative solution for preservation of known cells.
- The Anti-D does not detect the D VI variant.